

## Study on the Effect of Hydrosalpinx fluid on Sperm Motility

Tae-Cheol Kim, MD., Sang-Hoon Lee, MD.,

Dong-Ho Kim MD., Do-Hwan Bae, MD., Min Hur, MD.

Department of Obstetrics and Gynecology, College of Medicine, Chung-Ang University,  
Seoul, Korea

### =Abstract=

This study was performed to identify the effect of the hydrosalpinx fluid on sperm motility. It has been reported that the patients with hydrosalpinx show the outstandingly lower success rate than other patients having infertility by different factors. It is unclear that the cause of it is influenced by hydrosalpinx fluid directly or by secondary chronic inflammation of endometrium. We wanted to know if the hydrosalpinx fluid influences sperm motility parameters directly such that it is related to the development of infertility. Therefore, using computer assisted semen analyzer (CASA), we observed, from February to July, 1997, how sperm motility, sperm progressive motility, sperm curvilinear velocity, sperm lateral head displacement, sperm straightness and sperm linearity change after treating normal sperm with hydrosalpinx fluid to evaluate sperm function on infertility. The result was that the study group (n=32) has the tendency to differ from the control group (n=32) on sperm motility, progressive motility, curvilinear velocity, lateral head displacement, straightness and linearity. We concluded that the hydrosalpinx fluid, with varying degree, directly has the harmful effects on sperm motility parameters, that is, curvilinear velocity, lateral head displacement and linearity of sperm which are related to the hyperactivation, hence decreased capacitation.

Key words : hydrosalpinx fluid, sperm motility, IVF-embryo transfer

가 .

, Andersen (1996) 가

, , , cytokines, prostaglandins, leukotrienes, catecholamines 가

가 (Mukherjee et al., 1996 ).

(Denise et al., 1998),

가 . v 3 integrin 가 (Lessey et al, 1994). 가

(computer assisted sperm analyzer, CASA) 가 가 .

가 (Boyers et al., 1989 ; Mortimer., 1990). 가 (curvilinear velocity, VCL), (linear velocity), (average path velocity, VAP), (amplitude of lateral head displacement, ALH), (beat cross frequency, BCF), (straightness, STR) (linearity, LIN) .

가 (Burkman, 1984 ; Mortimer et al., 1990). Yanagimachi(1969) .

가 , (Domott and Suarez, 1992 ; Suarez and Dai, 1992). Burkman(1986) 가

가 .

가 .

1997 2 1997 7 , , 가 32 가 (HAM's F10 media)

600x g 10 37 ° c 0.22 μ m  
 filter  
 2-3 30-40  
 (WHO criteria,1987) , 80 x 10<sup>6</sup> sperm/ml , 30 50%  
 , 50% 0.5ml  
 2ml Ham 's 5% CO<sub>2</sub> 37 ° C 90  
 1000rpm 10  
 Hamilton - Thorn  
 가 100X10<sup>6</sup>/ml 1:1 5% CO<sub>2</sub> 37  
 1 , 5 , 24 , , , , ,

### 1. (motility)

1 (study group) 61.1 ±  
 12.4%, (control) 78.2 ± 7.4%  
 (p<0.05). 5 58.3 ±  
 13.2%, 76.2 ± 9.4% (P<0.01), 24  
 54.2 ± 12.2%, 73.2 ± 9.2% (P<0.01).(Table1)

Table1. Comparison of sperm motility(%) during culture in media with hydrosalpinx fluid(study group) and without hydrosalpinx fluid.(control group)

| incubation time(hour) | study group (%) (n=32) | control group (%) (n=32) |
|-----------------------|------------------------|--------------------------|
| 1                     | 66.1 ± 12.4*           | 78.2 ± 7.4               |
| 5                     | 58.3 ± 13.2**          | 76.2 ± 9.4               |
| 24                    | 54.2 ± 12.2**          | 73.2 ± 9.2               |

\*P<0.05

\*\*P<0.01

### 2. (progressive motility)

1 52.2 ± 11.2%, 61.2 ± 12.3%  
 가 (P<0.05), 5 42.3 ± 10.4%,  
 53.4 ± 12.8% (P<0.01) . 24

43.2 ± 9.8% (42.9 ± 9.2%)  
 .(Table 2)

Table 2. Comparison of sperm progressive motility(%) between study and control group.

| incubation time(hour) | study group (%) (n=32) | control group (%) (n=32) |
|-----------------------|------------------------|--------------------------|
| 1                     | 52.2 ± 11.2*           | 61.2 ± 12.3              |
| 5                     | 42.3 ± 10.4**          | 53.4 ± 12.8              |
| 24                    | 43.2 ± 9.8             | 42.2 ± 9.2               |

\*P<0.05

\*\*P<0.01

### 3. (curvilinear velocity)

1 38.2 ± 17.2um/ sec, 42.3 ± 12.4um/ sec  
 (P<0.05), 5 27.2 ± 12.6um/ sec,  
 36.2 ± 8.4um/ sec . 24  
 48.2 ± 10.2um/ sec, 63.4 ± 12.3um/ sec  
 (P<0.01).(Table 3)

Table 3. Comparison of sperm curvilinear velocity(um/sec) between study and control group

| incubation time(hour) | study group (um/sec)(n=32) | control group (um/sec)(n=32) |
|-----------------------|----------------------------|------------------------------|
| 1                     | 38.2 ± 17.2*               | 42.3 ± 12.4                  |
| 5                     | 27.2 ± 12.6                | 36.2 ± 8.4                   |
| 24                    | 48.2 ± 10.2**              | 63.4 ± 12.3                  |

\*P<0.05

\*\*P<0.01

### 4. (lateral head displacement, ALH)

1 1.82 ± 0.72um 2.31 ± 0.93um  
 . 5 2.47 ± 0.43um  
 2.74 ± 0.75um . 24  
 4.61 ± 0.31um 5.23 ± 0.36um  
 (P<0.05).(Table 4)

Table 4. Comparison of sperm ALH(um) between study and control group

| incubation time(hour) | study group (um) (n=32) | control group (um) (n=32) |
|-----------------------|-------------------------|---------------------------|
| 1                     | 1.82 ± 0.72             | 2.31 ± 0.93               |
| 5                     | 2.47 ± 0.43             | 2.74 ± 0.75               |
| 24                    | 4.61 ± 0.31*            | 5.23 ± 0.36               |

\*P<0.05

### 5. (straightness)

1 16.8 ± 2.8%, 17.4 ± 3.4%  
. 5 20.4 ± 3.2% 22.4 ± 6.2%  
. 24 15.4 ± 2.8%,  
19.2 ± 3.2% 가 (P<0.05).(Table 5)

Table 5. comparison of sperm straightness(%) between study and control group

| incubation time(hour) | study group (%) (n=32) | control group (%) (n=32) |
|-----------------------|------------------------|--------------------------|
| 1                     | 16.8 ± 2.8             | 17.4 ± 3.4               |
| 5                     | 20.4 ± 3.2             | 22.4 ± 6.2               |
| 24                    | 15.4 ± 2.8*            | 19.2 ± 3.2               |

\*P<0.05

### 6. (linearity)

1 23.5 ± 6.2%, 22.3 ± 4.8%  
. 5 22.3 ± 4.8%, 20.3 ± 6.2%  
. 24 20.7 ±  
5.3% 17.4 ± 3.2% (P<0.05).(Table 6)

Table 6. Comparison of sperm linearity(%) between control and study group

| incubation time(hour) | study group (%) (n=32) | control group (%) (n=32) |
|-----------------------|------------------------|--------------------------|
| 1                     | 23.5 ± 6.2             | 22.3 ± 4.8               |
| 5                     | 22.3 ± 4.8             | 20.3 ± 6.2               |
| 24                    | 20.7 ± 5.3*            | 17.4 ± 3.2               |

\*P<0.05

20%  
(Fergusson., 1982).  
70%  
50%  
가  
(Tan et al., 1992), (Steer et al., 1992),  
(Gonen et al., 1989)가  
가 (Lejeune et al., 1991).  
1/3  
(Vandromme et al., 1993 ; Sims et al., 1993). Katz (1996)  
(Andersen et al., 1996 ; Kassabji  
et al., 1994 ; Strandell et al., 1994 ; Vandromme et al., 1995), Robert Weiner  
가  
가 (Strandell, 1994 ; Anderson et al., 1996 ;  
Kassabji et al., 1994 ; Vandromme et al., 1995 ; Katz et al., 1996).  
Denise (1998)  
가 . Andersen (1996)  
. Kassabji (1994)  
, Robert Weiner (1997)  
가 (inflammatory hydrosalpinx)  
, (ongoing pregnancy rate) (Andersen et al., 1996 ;  
Kassabji et al 1994 ; Strandell et al 1994 ; Vandromme et al 1995 ; Bazer et al 1997)  
(overall success rate) (Shelton et al 1996 ; Katz et al  
1996 ; Bazer et al 1997). , , , , baby take home rate  
가

가 .

가 , 가 .

가 가 (David et al., 1969).

(Hunter, 1980).

가 가 가 가 가 (David A et al., 1969).

(embryo culture media) 가 (Mukherjee et al., 1996)

가 가 가 (Hill et al, 1987).

가

cytokines, prostaglandins, leukotrienes, catecholamines (David et al 1969 ; Owen et al, 1992),

가

(Strandell et al., 1994) (recruitment phase)

(Mansour et al., 1991), (Robert et al., 1997). - dysynchrony (Meyer et al., 1997)

v 3 integrin ,

(Lessey et al., 1994 ; Simon et al., 1995 ; Vanessa, 1997)

integrin (Meyer et al., 1997). v 3 integrin

가 (Lessey et al., 1994).

Vanessa (1997)

가

가 (Vanessa, 1997).

(O'Herlihy et al., 1981)

puncture toxic agent가

(Robert et al., 1997).

potassium bicarbonate, 가 (Garcia et al., 1969 ; Lippes et al., 1996) ( Lippes et al., 1996).

( Mansour et al, 1991 ; Andersen et al,1996 ; Kassabji et al 1994 ; Strandell et al 1994 ; Vandromme 1995 ; Bazer et al 1997)

C. trachomatis 가(antibody titer)가 가  
IVF cycle 10 doxycycline 100mg  
가 (Sharara et al., 1996).

neosalpingotomy, proximal tubal occlusion  
가  
가 (Danis et

al., 1998). Puttermans and Brosens(1996)  
(heterogeneous) 가

microsurgical  
salpingoneostomy . Puttermans (1996)

가 Andersen (1996)  
가

Andersen (1996)

가  
(inherent pathology of the disease  
Fallopian tube) IVF-ET  
(Denise et al., 1998).

(acute-phase) 가 (Mansour et al., 1991),  
1994), cytokine, prostaglandins, leukotrienes, inflammatory compounds  
가 가 (Grifo et  
al., 1989) integrin expression (Lessey et al., 1994), C. trachomatis  
(Sharara et al., 1996), biochemical imbalance

가 가 가 ,

가 , , ,

가 (Burkman., 1984 ; Mortimer et al., 1990). percoll swim  
up

가 , , 가 ( , 1996).

Yanagimachi(1969)

가 (capacitation) (Yanagimachi., 1969).

(acosome reaction)

가 , (Suarez and Dai., 1992). Burkman(1986)

가 1991 Burkman's criteria(linearity 65%, velocity 100um/ s, maxALH 7.5um)

가

1997 2 1997 7 HSG 32 (laparoscopy)

1. 1 (study group) 61.1 ± 12.4%, (control) 78.2 ± 7.4% (p<0.05). 5

58.3 ± 13.2%, 76.2 ± 9.4% (P<0.01), 24

54.2 ± 12.2%, 73.2 ± 9.2%

(P<0.01).

2. 1 52.2 ± 11.2%, 61.2 ± 12.3% 가 (P<0.05), 5 42.3 ± 10.4%, (P<0.01) . 24

53.4 ± 12.8% 43.2 ± 9.8% (42.9 ± 9.2%)

3. 1 38.2 ± 17.2um/ sec, 42.3 ± 12.4um/ sec (P<0.05), 5 27.2 ± 12.6um/ sec, 36.2 ± 8.4um/ sec . 24 48.2 ± 10.2um/ sec, 63.4 ± 12.3um/ sec (P<0.01).

4. 1 1.82 ± 0.72um 2.31 ± 0.93um . 5 2.47 ±



- Fergusson ILK. Laparoscopic investigation of tubal infertility. In : Chamberlain G. Winston R. eds. Tubal infertility : diagnosis and treatment. Oxford : Blackwell Sci Publ : 1982 ;30-46.
- Garcia CR, Czernobilsky B. Human hydrosalpinx. Am J Obstet Gynecol 1969 ; 105 : 400-10.
- Gonen Y, Casper RF, Jacobson W, Blankier J. Endometrial thickness and growth during ovarian stimulation: a possible predictor of implantation in vitro fertilization. Fertil Steril 1989 ; 52; 446-450.
- Grifo JA, Jeremias J, Ledger WJ, et al. Interferon gamma in the pathogenesis of pelvic inflammatory disease. Am J Obstet Gynecol 1989 ; 160 : 26.
- Hill JA, Hamiovici F, Politch JA, Anderson DJ. Effects of soluble products of activated lymphocytes and macrophages(lymphokines and monokines) on human sperm motion parameters. Fertil Steril 1987 ; 47 : 460.
- Hunter RHF. Physiology and technology of reproduction in female domestic animal. London : Academic Press. 1980 ; 748-884
- Kassabji M, Sims JA, Butler L, Muasher SJ. Reduced pregnancy outcome in patient with unilateral or bilateral hydrosalpinx after in vitro fertilization. Eur J Obstet Gynecol Reprod Biol 1994 ; 56:129-32
- Katz E, Akman M, Damewood MD, Garcia JE. Deleterious effect of the presence of hydrosalpinx on implantation and pregnancy rates with in vitro fertilization. Fertil Steril 1996;66:122-5
- Lejeune B. Vandromme J. Delvigne A. Puissant F. Van Rysselbege M, Barlow B, Leroy F. Lower implantation rate in IVF tubal indication(abstract). Presented at the 7th World Congress on In Vitro Fertilization and assisted Procreation. Paris, France, June 30-July 3, 1991
- Lessey BA, Castelbaum AJ, Riben M, Howarth J, Turek R, Meyer WR. Effect of hydrosalpinges on markers of uterine receptivity and success in IVF (abstract). 50th Annual Meeting of the American Society for for Reproductive Medicine.San Antonio, TX:American Society for Reproductive Medicine, 1994.
- Lippes J, Wagh PV. Human oviductal fluid (hOF) proteins. IV.Evidence for HOF proteins binding to human sperm Fertil Steril 1996;66:851-3
- Mansour RT, Aboughar MA, Serour GI, Riad R. Fluid accumulation of the uterine cavity before embryo transfer. Nippon Sanka Fujinka Gakkai Zasshi 1991 ; 43 : 1491-1498
- Meyer W, Castelbaum A, Somkuti S.(1997) Hydrosalpinges adversely affect markers of endometrial receptivity. Hum. Repro., 2, 1393-1398.
- Mortimer D. Objective analysis of sperm motility and kinematics. In : Keel BA & Webster BW eds. Handbook of the laboratory diagnosis and treatment of infertility. Florida : CRC press. iNC 1990 : 97.
- Mukherjee T, Copperman AB, McCaffrey C, Cook CA, Bustillo M, Obasaju MF.Hydrosalpinx fluid has embryotoxic effects on murine embryogenesis:a case for prophylactic salpingectomy.Fertil Steril 1996;66:851-3
- O'Herlihy C, DeCrespigny LJCh, Robinson H, Hoult I : The role of ultrasound in vitro fertilization program.(Abstr.) Fertil Steril 35 ; 235. 1981.

- Owen C, Kannisto P, Liedberg F. In: Sjoberg NO, Berger L, Janson PO, editors. Local regulation of ovarian function. Carnforth, UK: Parthenon, 1992 ; 149-70.
- Puttermans PJ, Brosens IA. Preventive salpingectomy of hydrosalpingectomy prior to IVF. Salpingectomy improves in vitro fertilization outcome in patient with a hydrosalpinx: blind timization of the fallopian tube. Hum Reprod 1996; 11: 20-81.
- Robert weiner, Lindhard A, Loft AI., The infertile patient with hydrosalpinges-IVF with or Without salpingectomy? Hum. Reprod., 1997 ; 11 : 2081-2084.
- Sharara FI, Scott RT, Marut EL, Queenan Jr. JT. In-vitro fertilization outcome in women with hydrosalpinx. Human Reprod 1996 ; 11 : 526-530
- Shelton KE, Butler L, Toner JP, Oehninger S, Muasher SJ. Salpingectomy improves the pregnancy rate in-vitro fertilization patients with hydrosalpinx.Hum Reprod 1996;11:523-5.
- Simon C, Mercader A, Portoles E, Frances A. Pellicer systeme interleukine-1 au cours de limplantation danpece humaine. Contracept. Fertil Sex 1995 ; 63 : 535-42.
- Sims JA, Jones D, Butler L, Muasher SJ. Effect of hydrosalpinx on outcome in vitro fertilization(IVF) (abstract). The American Fertility Society Meeting. Montreal, Canada. 1993 p.95.
- Steer CV, Mills CL, Tan SL, Cambell S, Edward RG. The cumulative enbryo score : a predictive embryo scoring technique to select the optimal number of embryos to transfer in an in vitro fertilization and embryo transfer program. Hum. Reprodm., 1992; 7 : 117-119.
- Strandell A, Wadenstrom U, Nilsson L. Hamberger L. Hydrosalpinx reduces in-vitro fertilization/embryo transfer pregnancy rates. Human reproduction 1994 ; 9 : 861-863.
- Suarez SS, Dai XB. Hyperactivation enhances mouse sperm capacity penetrating viscoelastic media. Biol Repro 1992 ; 46 : 686.
- Tan SE, Royston P, Cambell S, Jacobs HS, Betts J, Mason B, Edwards RG. Cumulative conception and livebirth rates after in-vitro fertilization. Lancet, 1992; 339 : 1390-1394.
- Vandromme J. Chasse E, Lejeune B. Hydrosalpinx in vitro fertilization : an unfavourable prognostic feature(Abstr).IVF Contact Group of Belgium Fund fr Medical Research, Brussell, March 6 ; 1993.
- Vandromme J, Chasse E, Leune B, Van Rysselberge M, Delvigne A, Leroy F. Hydrosalpinges in-vitro fertilization: an unfavourable prognostic feature. Hum Reprod 1995;10:576-9
- Vanessa J. Rawe, Mary G. Compton, M.A.S., Jiaen Liu, Jairo E. Garcia, Stefanie Shaffer, Eugene Katz. Effect of human hydrosalpinx fluid on murine embryo development and implantation. Fertil Steril. 1997 ; 68 ; 668-670
- Yanagimachi R. In vitro capacitation of hamster spermatozoa by follicular fluid. J Repro Fertil 1969 ; 18 : 275.

